

Urine Cell-Free Circulating and Viral Nucleic Acid Purification Kit

Product # 60000

Product Insert

Introduction

Recent evidence indicates that cell-free circulating (cfc) nucleic acids such as DNA and RNA, including exosomal RNA, in urine contains valuable information for the discovery of biomarkers that can help with early detection of certain cancer types, for monitoring the disease status as well as for the detection fetal DNA. Moreover, urine has been widely used for the detection of viral infections, including HIV, HCV and HBV. The advantage for using urine as a source for cancer biomarkers is that it can be acquired in large quantities without using invasive procedures, and repeated sampling from the same individual is applicable, which facilitates longitudinal studies. There are many advantages favouring the use of urinary nucleic acids for cancer biomarker discovery over blood, tissue samples or other bodily fluids, including: (1) urine is non-infectious for HIV and less infectious for many other pathogens; (2) the profile of urinary nucleic acids is similar to that in plasma or serum; (3) nucleic acid purification from urine is technically much easier because of its low protein concentration (1000-fold lower than blood).

Exosomes are 40 - 100 nm membrane vesicles, which are secreted by most cell types. Exosomes can be found in saliva, blood, urine, amniotic fluid and malignant ascitic fluids, among other biological fluids. Evidence has been accumulating recently that these vesicles act as cellular messengers, conveying information to distant cells and tissues within the body. The exosomes contain cell-specific proteins, lipids and RNAs, which are transported to other cells, where they can alter function and/or physiology. Recent work has demonstrated the presence of distinct subsets of microRNAs within exosomes which depend upon the tumour cell type from which they are secreted. For this reason exosomal RNAs may serve as biomarkers for various diseases including cancer. Cell-free mitochondrial DNA (cfmtDNA) is also under investigation for its clinical significance. In addition, cell-free fetal DNA has been widely used as a non-invasive method for prenatal diagnosis including early identification of fetal sex, genetic studies for families at high risk for inherited genetic disorders, screening for Rhesus factor, screening for aneuploidy and identification of preeclampsia.

Norgen's Urine Cell-Free Circulating and Viral Nucleic Acid Purification Kits provide a fast, reliable reproducible and simple procedure for isolating total circulating nucleic acids (DNA and RNA) from various urine inputs ranging from 250 μ L and up to 10 mL, with various kit formats addressing different urine input volumes. Purification is based on spin column chromatography that uses Norgen's proprietary resin separation matrix. The kit is designed to isolate all sizes of cfc-DNA and circulating RNA, including microRNA, as well as all sizes of exosomal RNA. Norgen's Urine Cell-Free Circulating and Viral Nucleic Acid Purification Kits provides a clear advantage over other available kits in that they do not require phenol/chloroform or any protease treatments. Moreover, the kit allows the user to elute into a flexible elution volume ranging from 50 μ L to 100 μ L. The purified nucleic Acid is of the highest integrity, and can be used in any downstream applications.

Kit Components

Component	Midi Kit (Cat.#60000)
Number of Preps	20 preps
Binding Solution K	75 mL
Lysis Buffer A	20 mL
Wash Solution A	18 mL
Elution Buffer F	6 mL
Mini Spin Columns	20
Midi Spin Column	20
Collection Tubes	20
Elution tubes (1.7 mL)	20
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These kits are suitable for the isolation of total nucleic acid (RNA and DNA) from fresh urine samples, frozen urine samples or urine samples collected on any urine preservative.

It has been found that urine samples stored at -70°C, -20°C or at 4°C will develop some precipitation due to the aggregation of some of the highly abundant proteins in urine. Eliminating these precipitates using centrifugation or filtration may cause the loss of some of the cfc protein-bound RNA. Furthermore, these precipitates may affect the quality of the purified nucleic acid. We recommend the use of Norgen's Urine Preservative when collecting urine samples. Norgen's Urine Preservative is designed for the preservation of nucleic acids and proteins in fresh urine samples at ambient temperatures, therefore no protein precipitation will occur and the purified nucleic acids will be of a higher quality. The components of the Urine Preservative allow samples to be stored for over 2 years at room temperature with no detected degradation of urine DNA, RNA or proteins. Norgen's Urine Preservative is available as a liquid format in Norgen's Urine Preservative Single Dose Ampules, as well as in a dried format in Norgen's Urine Collection and Preservation Tubes

Specifications

Kit Specifications	
Sample Type	Fresh or frozen urine
Sample volume Range	2 to 10 mL
Minimum Elution Volume	50 µL
Maximum Elution Volume	100 µL
Time to Complete 10 Purifications	40 - 45 minutes
Size of DNA Purified	≥ 50 bp
Size of RNA Purified	All sizes, including miRNA and small RNA (<200 nt)
Average Yields ¥	Variable depending on specimen

*** Please check page 6 for Average Urine Yields and Common RNA Quantification Methods**

Storage Conditions and Product Stability

All buffers should be kept tightly sealed and stored at room temperature. These kits are stable for 2 years after the date of shipment. It is recommended to warm Lysis Buffer A for 20 minutes at 60°C if any salt precipitation is observed.

Quality Control

In accordance with Norgen's Quality Management System, each lot of Norgen's Urine Cell-Free Circulating and Viral Nucleic Acid Purification Kits are tested against predetermined specifications to ensure consistent product quality.

Product Use Limitations

Norgen's Urine Cell-Free Circulating and Viral Nucleic Acid Purification Kits are designed for research purposes only. They are not intended for diagnostic use.

Product Warranty and Satisfaction Guarantee

NORGEN BIOTEK CORPORATION guarantees the performance of all products in the manner described in our product manual. The customer must determine the suitability of the product for its particular use.

Safety Information

Ensure that a suitable lab coat, disposable gloves and protective goggles are worn when working with chemicals. For more information, please consult the appropriate Material Safety Data Sheets (MSDSs). These are available as convenient PDF files online at www.norgenbiotek.com.

Binding Solution K and **Lysis Buffer A** contain guanidium salt, and should be handled with care. Guanidium salt forms highly reactive compounds when combined with bleach, thus care must be taken to properly dispose of any of these solutions.

If liquid containing these solutions is spilt, clean with suitable laboratory detergent and water. If the spilt liquid contains potentially infectious agents, clean the affected area first with laboratory detergent and water, and then with 1% (v/v) sodium hypochlorite.

CAUTION: DO NOT add bleach or acidic solutions directly to the sample-preparation waste.

Customer-Supplied Reagents and Equipments

- Benchtop microcentrifuge
- Swinging bucket centrifuges
- Vortexer
- Micropipettors
- 96 – 100% ethanol
- 100% Isopropanol
- β - Mercaptoethanol

Procedure

Important Notes

Working with RNA

RNases are very stable and robust enzymes that degrade RNA. Autoclaving solutions and glassware is not always sufficient to actively remove these enzymes. The first step when preparing to work with RNA is to create an RNase-free environment. The following precautions are recommended as your best defence against these enzymes.

- The RNA area should be located away from microbiological work stations
- Clean, disposable gloves should be worn at all times when handling reagents, samples, pipettes, disposable tubes, etc. It is recommended that gloves are changed frequently to avoid contamination
- There should be designated solutions, tips, tubes, lab coats, pipettes, etc. for RNA only
- All RNA solutions should be prepared using at least 0.05% DEPC-treated autoclaved water or molecular biology grade nuclease-free water
- Clean all surfaces with commercially available RNase decontamination solutions
- When working with purified RNA, ensure that they remain on ice during downstream applications

Notes Prior to Use

- All centrifugation steps are performed at room temperature.
- Ensure that centrifuge tubes used are capable of withstanding the centrifugal forces required.
- The provided spin columns are optimized to be used with a benchtop centrifuges and not to be used on a vacuum apparatus
- Most standard benchtop microcentrifuges will accommodate Norgen's Mini Spin Columns.
- Most standard swinging bucket centrifuges will accommodate Norgen's Midi and Maxi Spin Columns. Do not use a fixed-angle rotor
- Norgen's Midi and Maxi Spin Columns are centrifuged in 15 mL and 50 mL centrifuge tubes, respectively.
- Centrifuging Norgen's Spin columns at a speed higher than recommended may affect RNA yield.
- Centrifuging Norgen's Spin columns at a speed lower than recommended will not affect RNA yield. However, centrifugation at a lower speed may require longer time for the solutions to pass through the spin column
- When placing Norgen's Midi and Maxi Spin Columns into the swinging bucket centrifuge make sure that lids of the tubes are not tightly closed. Tightly closed lids may cause back pressure which may cause column clogging or disintegration.
- Ensure that all solutions are at room temperature prior to use.
- It is highly recommended to warm up **Lysis Buffer A** at 60°C for 20 minutes and mix well until the solutions become clear again if precipitates are present.
- Prepare a working concentration of the **Wash Solution A** by adding **42 mL** of 96 - 100% ethanol (provided by the user) to the supplied bottle containing **18mL** of concentrated Wash Solution A. This will give a final volume of **60 mL**. The label on the bottle has a box that may be checked to indicate that the ethanol has been added.
- The use of β -mercaptoethanol in lysis is highly recommended to isolate RNA for sensitive downstream applications. Add 10 μ L of β -mercaptoethanol (provided by the user) to each 1 mL of **Lysis Buffer A**.
- ***If any of the solutions do not go through the Spin Columns within the specified centrifugation time, spin for an additional 1-2 minutes until the solution completely passes through the Column. Do NOT exceed the centrifugation speed as this may affect RNA yield.***

Preparation of Cell-free Urine Sample

1. Collect and transfer 15-50 mL of urine into a conical tube and centrifuge at **200 x g (~1,000 RPM) for 10 minutes** to remove urine exfoliated cells and debris. Decant cell-free urine into new 15-50 mL conical tube.
 2. Centrifuge the cell-free urine at **1,800 x g (~3,000 RPM) for 10 minutes** to remove any residual debris or bacterial cells.
 3. Transfer cell-free urine into a fresh 15-50 mL conical tube.
- **Cell-Free Urine is now ready for the purification of cell-free DNA**

Note: The procedure outlined below is for 10 mL inputs of urine. If processing a sample volume lower than 10 mL urine, simply bring the volume of your samples up to 10 mL using Nuclease-free water and proceed as outlined below.

1. Place 10 mL of cell-free urine in a 50 mL tube (provided by the user). Add 3 mL **Binding Solution K** and mix well by vortexing for 10 seconds.
2. Transfer 4.5 mL of the mixture from **Step 1** into a Midi Spin column assembled with one of the provided collection tubes. Centrifuge for **3 minutes at 1,000 x g (~2,200 RPM)**. Discard the flowthrough and reassemble the spin column with its collection tube.
3. Repeat **Step 2** two more times to transfer the remaining mixture into the Midi Spin column.
4. Apply 400 µL of **Lysis Buffer A** to the column and centrifuge for **3 minutes at 500 x g (~1,600 RPM)**. **Do Not Discard the flowthrough.**
5. Apply an additional 400 µL of **Lysis Buffer A** to the column and centrifuge for **3 minutes at 500 x g (~1,600 RPM)**. **Do Not Discard the flowthrough.**
6. Reload the 800 µL flowthrough from **Step 5** back to the column and let stand at room temperature for 20 minutes. Centrifuge for **3 minutes at 1,000 x g (~2,200 RPM)**. **Do Not Discard the flowthrough.**
7. To the flowthrough from **Step 6** add 400 µL **100% Isopropanol** and mix well by pipetting.
8. Transfer 600 µL of the mixture from **Step 7** to a Mini Spin column assembled with one of the provided collection tubes. Centrifuge for **2 minutes at 3,300 x g (~7,000 RPM)**. Discard the flowthrough and reassemble the spin column with its collection tube.
9. Repeat **Step 8** one more time to transfer the remaining mixture into the Mini Spin column.

Optional Step: If only RNA is the primary interest, an optional **On-Column DNA Removal Protocol** is provided in **Appendix A** for the maximum removal of DNA that may affect sensitive downstream applications. This step should be performed at this point in the protocol

10. Apply 600 µL of **Wash Solution A** to the column and centrifuge for **1 minute at 3,300 x g (~7,000 RPM)**. Discard the flowthrough and reassemble the spin column with its collection tube.
11. Repeat **Step 10** one more time, for a total of two washes.
12. Spin the column, empty, for **2 minutes at 14,000 x g (~14,000 RPM)**. Discard the collection tube.
13. Transfer the spin column to a fresh 1.7 mL Elution tube. Apply 50 µL of **Elution Buffer F** to the column and let stand at room temperature for 2 minutes. Centrifuge for **1 minute at 200 x g (~2,000 RPM)**, followed by **2 minutes at 5,200 x g (~8,000 RPM)**.
14. For maximum recovery, transfer the eluted buffer back to the column and let stand at room temperature for 2 minutes. Centrifuge for **1 minute at 400 x g (~2,000 RPM)**, followed by **2 minutes at 5,800 x g (~8,000 RPM)**.

- **Urine nucleic acid is ready for the downstream application of your choice. For an explanation of expected yields and recommendations for quantification of the nucleic acid, please refer to Appendix B**

Appendix A

Protocol for Optional On-Column DNA Removal

Norgen's Urine Cell-Free Circulating and Viral Nucleic Acid Purification Kits isolate total nucleic acid (RNA and DNA) in one elution. However, an optional protocol is provided below for the maximum removal of DNA that may affect sensitive downstream applications if your primary interest is cell-free circulating RNA. It is recommended that Norgen's RNase-Free DNase I Kit (Product # 25710) be used for this step.

1. For every on-column reaction to be performed, prepare a mix of 15 μL of **DNase I** and 100 μL of **Enzyme Incubation Buffer** using Norgen's RNase-Free DNase I Kit (Product # 25710). Mix gently by inverting the tube 5-10 times. **DO NOT VORTEX.**
Note: If using an alternative DNase I, prepare a working stock of 0.25 Kunitz unit/ μL RNase-free DNase I solution according to the manufacturer's instructions. A 100 μL aliquot is required for each column to be treated.
2. Perform the procedure up to **Step 9.**
3. Apply 400 μL of **Wash Solution A** to the column and centrifuge for **30 seconds at 6,000 RPM.** Discard the flowthrough and reassemble the spin column with its collection tube.
4. Apply 50 μL of the **RNase-free DNase I** solution prepared in **Step 1** to the column and centrifuge at **8,000 x g (~10,000 RPM)** for 1 minute.
Note: Ensure that the entire DNase I solution passes through the column. If needed, spin at **13,000 x g (~14,000 RPM)** for an additional minute.
5. After the centrifugation in **Step 4**, pipette the flowthrough that is present in the collection tube back onto the top of the column.
Note: Ensure **Step 5** is performed in order to ensure maximum DNase activity and to obtain maximum yields of RNA, in particular for small RNA species.
6. Incubate the column assembly at 25 - 30°C for 15 minutes.
7. Without any further centrifugation, proceed directly **to the second wash step** in **Step 11.**

Appendix B

Cell-Free Circulating Nucleic Acid Yield

Urine total Nucleic Acid, like RNA and DNA in other cell-free bodily fluids, is normally found in very low amounts (1 - 100 pg/ μL), therefore measuring cell-free RNA or DNA concentration using common quantification methods is very difficult and challenging. Typical yields of urine Nucleic Acid vary significantly from sample to sample. Variability is also observed between samples collected from the same donor at different times during the day and therefore there is no absolute yield for RNA purified from bodily fluids including urine. Cell-free circulating RNA yield varies depending on a number of factors including age, sex, diet, exercise and most importantly the health status of the donor.

Below is a list of the most common RNA and DNA quantification methods, as well as the limit of detection for each of these methods. **Unfortunately, none of these methods can be used reliably for measuring the concentration of the purified nucleic acid from urine unless large urine volumes have been processed.** This would only be applicable if urine contains the maximum amount of nucleic acid that can fit within the specification range of these quantification tools. It should be noted that the specifications outlined below are based on measuring a pure RNA/DNA, which will not be the case for the nucleic acid purified from urine. Urine nucleic acid is short and fragmented, and is usually present in less than 1000 bp. Purified urine nucleic acid

usually contains traces of proteins which will interfere with most quantification methods, leading to the overestimation of the purified nucleic acid concentration. Therefore purified nucleic acid contaminated with more proteins will be presented at a higher concentration as compared to nucleic acid purified with less protein contaminants, which in this case will depend on the method used for urine nucleic acid purification. **The only reliable method that can assess the quality and the relative quantity of the purified urine nucleic acid is RT-qPCR (RNA) or qPCR (DNA) amplification of a standard RNA or DNA using a small amplicon such as the 5S rRNA housekeeping gene.**

Common Nucleic Acid Quantification Methods

1) Bioanalyzer RNA Quantification Kits

	RNA 6000 Nano Kit		RNA 6000 Pico Kit		Small RNA kit
	Total RNA	mRNA	Total RNA	mRNA	Total RNA
Quantitative range	25 - 500 ng/μL	25 - 250 ng/μL	----	----	50-2000 pg/μL
Qualitative range	5 - 500 ng/μL	5 - 250 ng/μL	50 - 5000 pg/μL	250 - 5000 pg/μL	50-2000 pg/μL
Quantitation accuracy	20% CV	20% CV	30% CV	----	-----

2) Bioanalyzer DNA Quantification Kits

	DNA 1000 Kit	DNA 7500 Kit	DNA 12000 Kit	High Sensitivity DNA Kit
Size Range	25–1000 bp	100–7500 bp	100–12000 bp	50-7000 bp
Quantitation accuracy	20% CV*	20% CV*	25% CV*	20% CV
Quantitative range	0.5-50 ng/μL	0.5-50 ng/μL	0.5-50 ng/μL	5-500 pg/μL

3) NanoDrop 2000

- Detection Limit: 2 ng/μl (dsDNA)

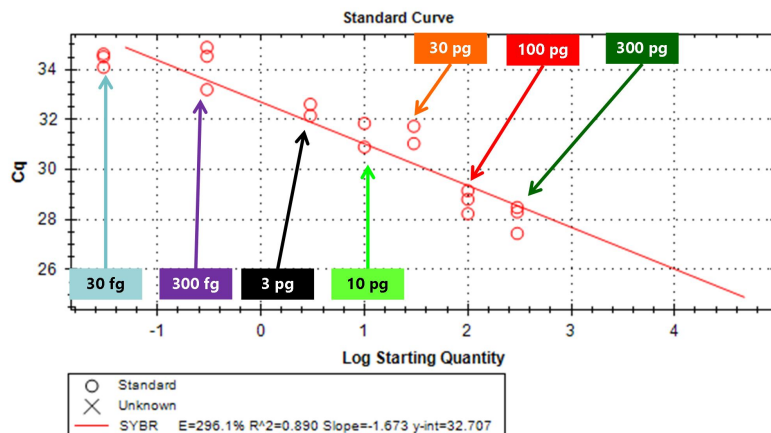
4) Quant-iT™ RiboGreen® RNA Assay Kit

- Quantitation Range: 1-200 ng

5) Quant-iT™ Pico Green® dsDNA Assay Kit

- Detection Limit: 25 pg/mL

6) qPCR Standard Curve (generated using Norgen's Low Abundance DNA Quantification Kit, Cat# 57200)



Frequently Asked Questions

1. What if a variable speed centrifuge is not available?

- A fixed speed centrifuge can be used, however reduced yields may be observed.

2. At what temperature should I centrifuge my samples?

- All centrifugation steps are performed at room temperature. Centrifugation at 4 °C will not adversely affect kit performance.

3. What if I added more or less of the specified reagents' volume?

- Adding more or less than the specified volumes may reduce both the quality and the quantity of the purified nucleic acids. Eluting your nucleic acids in high volumes will increase the yield but will lower the concentration. Eluting in small volumes will increase the concentration but will lower the overall yield.

4. What if I forgot to do a dry spin before my final elution step?

- Your purified nucleic acids will be contaminated with the Wash Solution A. This may reduce the quality of your purified nucleic acid and will interfere with your downstream applications.

5. Can I perform a second elution?

- Yes, but it is recommended that the 2nd elution be in a smaller volume (50% of 1st Elution). It is also recommended to perform the 2nd elution into a separate elution tube to avoid diluting the 1st elution.

6. Why do my samples show low nucleic acid yield?

- Urine samples contain very little nucleic acid. This varies from individual to individual based on numerous variables. In order to increase the yield, the amount of urine input could be increased.

7. Why do the A260:280 ratio of the purified nucleic acid is lower than 2.0?

- Most of the Free-Circulating urine nucleic acid is short nucleic acid fragments. The A260:280 ratio is normally between 1 – 1.6. This low A260:280 ratio will not affect any downstream application.

8. Why does my isolated RNA not perform well in downstream applications?

- If a different Elution Buffer was used other than the one provided in the kit, the buffer should be checked for any components that may interfere with the application. Common components that are known to interfere are high salts (including EDTA), detergents and other denaturants. Check the compatibility of your elution buffer with the intended use.

9. Do I need to do a DNase treatment for my nucleic acid Elution?

- You may need to do a DNase treatment to your isolated urine miRNA. It is recommended to use Norgen's RNase-Free DNase I Kit (Cat# 25710). Also please refer to the protocol for optional on-column DNA removal outlined in **Page 6**.

Technical Support

Contact our Technical Support Team between the hours of 9:00 and 5:30 (Eastern Standard Time) at (905) 227-8848 or Toll Free at 1-866-667-4362.

Technical support can also be obtained from our website (www.norgenbiotek.com) or through email at support@norgenbiotek.com.

Norgen's purification technology is patented and/or patent pending. See www.norgenbiotek.com/patents

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